SHORT REPORTS

ISOLATION OF PARASORBIC ACID FROM THE CRANBERRY PLANT, VACCINIUM MACROCARPON

J. H. CARDELLINA II and J. MEINWALD

Department of Chemistry, Cornell University, Ithaca, NY 14853, U.S.A.

(Received 6 Febuary 1980)

Key Word Index—Vaccinium macrocarpon; Ericaceae; cranberry; parasorbic acid; seed germination inhibition.

Abstract—Parasorbic acid, 5,6-dihydro-6(S)-methyl-2H-pyran-2-one, has been isolated from whole plant extracts of the cranberry, *Vaccinium macrocarpon*. This antibiotic lactone, which also inhibits seed germination, has been reported previously only from the fruit of the mountain ash, *Sorbus aucuparia*.

Parasorbicacid, 1, has been known for over a century [1] to be a constituent of the fruits of Sorbus aucuparia (Rosaceae). This unsaturated lactone has been shown to inhibit fungal growth [2] and seed germination [3, 4]. More recently, it has been reported [5] that 1 is derived from the glucoside 2 in Sorbus fruits. To our knowledge, neither 1 nor 2 has been reported from any natural source other than Sorbus.

In the course of an examination of cranberry plant extracts for alkaloids, we isolated parasorbic acid, 1, from a methanol-soluble fraction in 0.12% yield (based on plant dry wt). Since the plant extracts were exposed to acid during the solvent partitioning sequence which led to the isolation of 1, it is possible that 1 is produced by acid hydrolysis of 2 or a related glycoside. The ¹H NMR spectrum indicated the presence of eight protons—a methyl group, an allylic methylene, a methine on carbon bearing a heteroatom and two olefinic protons. Decoupling experiments led to partial structure 1a. The IR spectrum gave evidence for the presence of an ester or lactone and the absence of hydroxyl groups. The mass

spectrum showed a molecular ion at m/e 112; in conjunction with the IR spectral data, this suggested that the elements of CO_2 were missing from 1a. The identification of 1 was confirmed by comparison of the reported optical rotation [6] and UV [7], IR [7], MS [8] and ¹H NMR [9] spectra of parasorbic acid with those of our compound.

Inhibition of seed germination and plant growth by extracts of cranberry plants has recently been demonstrated (R. M. Devlin, personal communication). Since 1 is known to exhibit such inhibitory activity against a variety of plants [3, 4], it may well be responsible for the allelochemical activity in *V. macrocarpon* and, at the same time, offer some protection to the plant from fungal infection.

EXPERIMENTAL

¹H NMR spectra were obtained using a Varian EM-390 Spectrometer; chemical shifts are reported in δ units (ppm) relative to TMS ($\delta = 0$) as an internal standard in CDCl₃.

Isolation of 1. The aerial portion of Vaccinium macrocarpon was air-dried (382 g) and Soxhlet-extracted, first with petrol, then with CH₂Cl₂. The plant residue was then steeped in MeOH for 2 days. The MeOH extracts were reduced in vol. to 450 ml, diluted to 500 ml with 0.5 M HCl, and extracted with hexane; the acidic ag. MeOH phase was subsequently diluted to 75% MeOH with distilled H2O, extracted with CCl4, and then further diluted to 65% MeOH and extracted with CHCl₃. MeOH was removed from the aq. phase at red. pres.; the residual suspension was diluted to 500 ml with H2O and extracted with EtOAc. Finally, the aq. phase was made basic with K₂CO₃ and extracted with CHCl3. The CHCl3 extracts were dried and reduced, in vacuo, to give a pleasant-smelling brown oil, 890 mg. Sephadex LH-20 gel filtration of 65 mg of the oil, using CH₂Cl₂-MeOH (1:1), gave 32 mg of 1, a colorless mobile oil, $[\alpha]_D^{25} + 181^\circ$ (EtOH, c1.0) [lit. [6] $[\alpha]_D^{25} + 197^\circ$ (EtOH, c6.3)]; ¹H NMR: δ 6.95 (1 H, ddd, J = 10, 5, 3 Hz), 5.98 (1 H, dt, J = 10, 1.5 Hz), 4.57 (1 H, dq, J = 9, 6 Hz), 2.33 (2 overlapping 1H, ddd, J = 9, 3, -14 and J = 6, 5, 2200 Short Reports

-14 Hz), 1.45 (3 H, d, J=6 Hz); UV; $\lambda_{\rm max}^{\rm EIOH}$ 221 nm (ε 8100); IR (CHCl₃) cm⁻¹: 2960, 2920, 2900, 1730, 1635, 1465, 1455, 1400, 1365, 1310, 1265, 1125, 1110, 1055, 1000, 955, 855; MS m/e (rel. int): 112 (2), 97 (7), 68 (100), 40 (15), 39 (13).

Acknowledgements—We thank Dr. Karl H. Deubert of the University of Massachusetts Cranberry Experiment Station for providing the cranberry plants. Partial support of this research by the Schering Corporation is gratefully acknowledged.

REFERENCES

- 1. Hofmann, A. W. (1859) Annalen 111, 129.
- 2. Buston, H. W. and Roy, S. K. (1949) Arch. Biochem. 22, 1.

- Buston, H. W., Roy, S. K., Hatcher, E. S. J. and Rawes, M. R. (1949) Arch. Biochem. 22, 269.
- 4. Moewus, F. and Schader, E. (1951) Z. Naturforsch. 66, 112.
- Tschesche, R., Hoppe, H. J., Snatzke, G., Wulff, G. and Fehlhaber, H. W. (1971) Chem. Ber. 104, 1420.
- 6. Kuhn, R., and Jerchel, D. (1943) Chem Ber. 76, 413.
- Brunn, J., Dethloff, M. and Riebenstahl, H. (1977) Z. Phys. Chem. Leipzig 258, 209.
- 8. Crombie, L. and Firth, P. A. (1968) J. Chem. Soc. 2852.
- Jackman, L. M. and Sternhell, S. (1969) Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry, p. 188. Pergamon Press, Oxford.